K_{IR} channels function as electrical amplifiers in rat vascular smooth muscle

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Strong inward rectifying K^+ (K_{IR}) channels have been observed in vascular smooth muscle and can display negative slope conductance. In principle, this biophysical characteristic could enable K_{IR} channels to 'amplify' responses initiated by other K^+ conductances. To test this, we have characterized the diversity of smooth muscle K_{IR} properties in resistance arteries, confirmed the presence of negative slope conductance and then determined whether K_{IR} inhibition alters the responsiveness of middle cerebral, coronary septal and third-order mesenteric arteries to K^+ channel activators. Our initial characterization revealed that smooth muscle K_{IR} channels were highly expressed in cerebral and coronary, but not mesenteric arteries. These channels comprised K_{IR}2.1 and 2.2 subunits and electrophysiological recordings demonstrated that they display negative slope conductance. Computational modelling predicted that a K_{IR}-like current could amplify the hyperpolarization and dilatation initiated by a vascular K^+ conductance. This prediction was consistent with experimental observations which showed that 30 μM Ba^{2+} attenuated the ability of K^+ channel activators to dilate cerebral and coronary arteries. This attenuation was absent in mesenteric arteries where smooth muscle K_{IR} channels were poorly expressed. In summary, smooth muscle K_{IR} expression varies among resistance arteries and when channel are expressed, their negative slope conductance amplifies responses initiated by smooth muscle and endothelial K^+ conductances. These findings highlight the fact that the subtle biophysical properties of K_{IR} have a substantive, albeit indirect, role in enabling agonists to alter the electrical state of a multilayered artery.

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The magnitude and distribution of tissue blood flow is controlled by an integrated network of resistance arteries (Segal & Duling, 1986; Segal, 2000). Under dynamic conditions, tone within an arterial network is regulated by multiple stimuli initiated by changes in intraluminal pressure (Bayliss, 1902; Knot & Nelson, 1998), blood flow (Garcia-Roldan & Bevan, 1990; Koller & Kaley, 1991), neuronal activity (Brayden & Bevan, 1985; Si & Lee, 2002) and tissue metabolism (Harder *et al.* 1998; Filosa *et al.* 2006). Vasoactive stimuli influence arterial diameter by activating signal transduction pathways which control myofilament Ca^{2+} sensitivity (Somlyo & Somlyo, 2003) and/or cytosolic [Ca^{2+}] (Nelson *et al.* 1990; Nelson & Quayle, 1995). Cytosolic [Ca^{2+}] is, in turn, tightly coupled to membrane potential (V_M) and the graded influx of Ca^{2+} through voltage-operated Ca^{2+} channels (Nelson *et al.* 1990; Nelson & Quayle, 1995).

To initiate changes in smooth muscle V_M, vasoactive agents must directly regulate an ionic conductance. This regulation is frequently viewed in simple terms with stimuli activating transduction pathways that control kinases and phosphatases responsible for channel phosphorylation (Nelson *et al.* 1990; Nelson & Quayle, 1995). Although direct regulation is essential, this is often interpreted to suggest that only these currents alter the electrical and mechanical state of vascular smooth muscle. This view overlooks the potential contribution of ionic conductances whose voltage-dependent properties could in essence 'facilitate' or 'amplify' an electrical response initiated by another agonist-sensitive channel.

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Of particular interest is the strong inward rectifying K^+ (K_{IR}) current which over the physiological voltage range displays negative slope conductance. (Matsuda *et al.* 2003; Schram *et al.* 2003; Dhamoon *et al.* 2004). Briefly, negative slope conductance refers to an inherent ability of some K_{IR} channels to increase their activity as a cell hyperpolarizes (Nelson *et al.* 1990; Nelson & Quayle, 1995). Such an increase contrasts with the situation for other vascular K^+ channels and is dependent upon the relief of a voltage-dependent Mg^{2+}/polyamine block (Nelson & Quayle, 1995; Robertson *et al.* 1996; Matsuda *et al.* 2003). Negative slope conductance is typically observed in channels composed of K_{IR}2.1 and 2.2 subunits and these constituents are often expressed in vascular smooth muscle (Bradley *et al.* 1999; Karkanis *et al.* 2003; Wu *et al.* 2007). While smooth muscle K_{IR} channels could, theoretically, amplify arterial responses, the supporting experimental evidence remains limited.

To explore whether smooth muscle K_{IR} channels operate as electrical amplifiers, we characterized K_{IR} expression in resistance arteries, confirmed negative slope conductance and then determined whether channel inhibition influenced the responsiveness of middle cerebral, coronary septal and third-order mesenteric arteries to K^+ channel activators. Findings show that smooth muscle K_{IR} channels are differentially expressed among resistance arteries, and when present are composed of K_{IR}2.1 and 2.2 subunits displaying negative slope conductance. Computational and experimental analyses reveal that this biophysical property enables smooth muscle K_{IR} to augment the electrical and vasomotor responses initiated by other smooth muscle or endothelial K^+ channels. These findings demonstrate that subtle channel properties are important for the regulation of arterial responsiveness. They also indicate that an ionic conductance need not be directly regulated by an agonist to significantly alter the electrical and mechanical state of vascular smooth muscle.

Methods

Animal procedures

Animal procedures were approved by the Animal Care and Use Committee at the University of Calgary. Briefly, female Sprague–Dawley rats (10–12 weeks of age) were killed via carbon dioxide asphyxiation. The brain, heart and mesentery were carefully removed and placed in cold phosphate-buffered (pH 7.4) saline solution containing (mM): NaCl 138, KCl 3, Na_2HPO_4 10, NaH_2PO_4 2, glucose 5, $CaCl_2$ 0.1 and $MgSO_4$ 0.1. Middle cerebral, coronary septal and third-order mesenteric arteries were carefully dissected out of surrounding tissue and cut into 2 mm segments.

Vessel myography

Arterial segments were mounted in a customized arteriograph and superfused with warm (37°C) physiological salt solution (PSS; pH 7.4) containing (mM): NaCl 119, KCl 4.7, $NaHCO_3$ 20, KH_2PO_4 1.1, $MgSO_4$ 1.2, $CaCl_2$ 1.6 and glucose 10. With the exception of vessels used for experiments in Fig. 9, endothelial cells were removed from all vessels by passing air bubbles through the vessel lumen (2–4 min); successful removal was confirmed by the loss of acetylcholine- or bradykinin-induced dilatation. Arteries were equilibrated for 60 min and contractile responsiveness assessed by brief (~10 s) exposure of the tissue to 60 mM KCl. Following equilibration, arteries were maintained in a hyperpolarized (15 mmHg intravascular pressure) or depolarized (80 mmHg + 0.01–1 μM phenylephrine) state. Vessels were then exposed to one of four experimental protocols: (1) Ba^{2+} (30 μM) alone; (2) elevated extracellular $[K^+]$ (raised from 5.8 or 4.0 to 15.0 mM) ± Ba^{2+} or ouabain (100 nM); (3) P-1075 or pinacidil (0.01–10 μM, K_{ATP} channel openers) ± Ba^{2+}; or (4) superfused acetylcholine (0.1–10 μM) or intraluminal uridine triphosphate (UTP, 2 μM) ± Ba^{2+}. Control experiments confirmed that glybenclamide (10 μM) blocked the ability of cerebral ($n = 2$) and coronary ($n = 2$) arteries to dilate in response to P-1075 (1 μM) and pinacidil (1 μM), respectively. Arterial diameter was monitored using an automated edge detection system (IonOptix, Milton, MA, USA). Smooth muscle V_M was assessed by inserting a glass microelectrode backfilled with 1 M KCl (tip resistance, 120–150 MΩ) into the vessel wall. The criteria for successful cell impalement included: (1) a sharp negative V_M deflection upon entry; (2) a stable recording for at least 1 min following entry; and (3) a sharp return to baseline upon electrode removal.

Isolation of arterial smooth muscle cells

Smooth muscle cells from middle cerebral, coronary septal and third-order mesenteric arteries were enzymatically isolated as previously described (Welsh & Brayden, 2001; Luykenaar *et al.* 2004). Briefly, arterial segments were placed in an isolation medium (37°C, 10 min) containing (mM): NaCl 60, sodium glutamate 80, KCl 5, $MgCl_2$ 2, glucose 10 and Hepes 10 with 1 mg ml^{-1} albumin (pH 7.4). Vessels were then exposed to a two-step digestion process that involved: (1) a 10–20 min incubation in isolation media (37°C) containing 0.6 mg ml^{-1} papain and 1.8 mg ml^{-1} dithioerythritol; and (2) a 10–20 min incubation in isolation medium containing 100 μM Ca^{2+}, 0.7 mg ml^{-1} type F collagenase and 0.4 mg ml^{-1} type H collagenase. Following treatment, tissues were washed repeatedly with ice-cold isolation medium and triturated

with a fire-polished pipette. Liberated cells were stored in ice-cold isolation medium for use the same day.

Electrophysiology

Conventional patch-clamp electrophysiology was used to measure whole-cell currents in isolated smooth muscle cells. Briefly, recording electrodes (resistance, 4–7 MΩ) were fashioned from borosilicate glass, covered in sticky wax to reduce capacitance, and backfilled with solution containing (mM): NaCl 5, KCl 35, potassium gluconate 100, $CaCl_2$ 1, Hepes 10, EGTA 10, Tris-ATP 2.5 and GTP 0.2 (pH 7.2). This pipette was then gently lowered onto a cell and negative pressure applied to rupture the membrane. Cells with an input resistance greater than 10 GΩ were then voltage clamped (−60 mV) and equilibrated for 15 min in a bath solution containing (mM): NaCl 135, KCl 5, $MgCl_2$ 0.1, Hepes 10, glucose 5 and $CaCl_2$ 0.1 (pH 7.4). Following equilibration, bath $[K^+]$ was elevated from 5 to 60 mM via equimolar replacement of NaCl by KCl. K_{IR} activity was then assessed by ramping cells between −120 and 20 mV (0.047 mV ms^{-1}), and quantifying the component of the whole-cell current that was sensitive to 30 μM Ba^{2+}. Control experiments monitoring the glybenclamide-sensitive (10 μM; at −80 mV with 60 mM extracellular K^+) and the iberiotoxin-sensitive (100 nM; +20 mV) currents confirmed that the pipette solution minimized K_{ATP}-activated and Ca^{2+}-activated K^+ (K_{Ca}) channel activity, respectively. Whole-cell currents were recorded on an Axopatch 200B amplifier (Axon Instruments, Union City, CA, USA), filtered at 1 kHz, digitized at 5 kHz and stored on a computer for subsequent analysis with Clampfit 8.1 software. Cell capacitance ranged between 14 and 18 pF and was measured with the cancellation circuitry in the voltage-clamp amplifier. Cells that displayed a noticeable shift in capacitance (> 0.3 pF) during experiments were excluded from analysis. A 1 M NaCl–agar salt bridge between the reference electrode and the bath solution was used to minimize offset potentials (< 2 mV). All experiments were performed at room temperature (20–22°C).

Real-time PCR analysis of K_{IR} subtypes

Smooth muscle cells (∼200) isolated from middle cerebral, coronary septal and third-order mesenteric arteries were placed in RNase- and DNase-free collection tubes. Total RNA was extracted (RNeasy mini kit with DNAase treatment; Qiagen, Valencia, CA, USA) and first-strand cDNA synthesized using the Sensi-script RT kit (Qiagen) with oligo d(T) primer. To optimize reaction specificity, real-time PCR was initially performed with each primer set using rat brain cDNA, SYBR green (Bio-Rad, Hercules, CA, USA), and a range of annealing temperatures (52–62°C). Following melt-curve analysis, 1 μl each reaction product was placed on a DNA 500 LabChip and examined using a bioanalyser (Model 2100, Agilent Technologies). A second aliquot of product was electrophoresed on a 1.5% (wt/vol) agarose gel, extracted using a gel extraction kit (Qiagen), and sequenced at the University of Calgary Core DNA facility. Having determined an ideal annealing temperature, real-time PCR efficiency was determined for all primer sets, with serial dilutions of brain cDNA used as template. Reaction efficiencies were as follows: KIR2.1, 94.4%; KIR2.2, 94.9%; KIR2.3, 98.8%; KIR2.4, 98.0%; and β-actin, 90.1%. The optimal real-time PCR reaction consisted of a hot start (95°C for 3 min) followed by 40 cycles of 95°C for 15 s, 55.1°C for 30 s and 72°C for 30 s. Samples were then exposed to a final extension period at 72°C for 10 min. A melt curve analysis was performed on each reaction and the threshold cycle determined using software provided with the Bio-Rad iCycler. K_{IR} mRNA levels were standardized to β-actin. Forward (F) and reverse (R) primers were as follows: KIR2.1 (accession no. NM_017296) (F) 5′- AGAGGAAGAGGACAGTGAGAAC-3′, K_{IR}2.1 (R) 5′-TCGCCTGGTTGTGGAGATC-3′; K_{IR}2.2 (accession no. NM_053981) (F) 5′-GCAGCCTTTCTCTTCTCCA-TTGA-3′, K_{IR}2.2 (R) 5′-GACTGAGCCACCACCATGA AG-3′; K_{IR}2.3 (accession no. NM_053870) (F) 5′-CCTG-GACCGCATCTTCTTGG-3′, K_{IR}2.3 (R) 5′-CAGGAT-GACCACAATCTCAAAGTC-3′; K_{IR}2.4 (accession no. NM_170718) (F) 5′-ATGAGGTTGACTATCGACACTT-CC-3′, K_{IR}2.4 (R) 5′-GGGAGCCAGGAAAACTTGAC-TTA-3′; β-actin (accession no. NM_031144) (F) 5′TATGAGGGTTACGCGCTCCC3′ (R) 5′ACGCTC-GGTCAGGATCTTCA3′. All smooth muscle cell samples were screened for template and endothelial cell contamination as previously described (Wu *et al.* 2007).

Computational modelling

To strengthen our experimental approach, a computational model developed by Diep *et al.* (2005) predicted whether a smooth muscle K_{IR}-like current could facilitate arterial hyperpolarization. Computational theory and base parameters were similar to those of the original model (Diep *et al.* 2005) with two notable exceptions. First, three additional layers of smooth muscle cells were added to our virtual artery, which is consistent with ultrastructural findings (Fig. 10). Second, the non-linear resistor representing the smooth muscle ionic conductance was divided into two components, one for K_{IR} and the second for all other conductances. Based on the results of previous studies, we assumed that K_{IR} was minimally active at −40 mV (0 pA), maximally active at −60 mV (2.0 pA) and reversed at −80 mV (Wu *et al.* 2007). A sigmoidal and exponential function fixed the data points between −40 and −60 mV, and −60 and −80 mV, respectively. At 2.0 pA, peak outward current would

be close to or below the resolution limit of whole-cell patch-clamp electrophysiology. The non-linear resistor representing all other smooth muscle conductances was comparable to that of Diep *et al.* (2005) except that its relative magnitude (below −40 mV) was increased by 15%. Hyperpolarization was initiated by injecting 4.6 pA current into each smooth muscle cell for 500 ms. Voltage responses were subsequently monitored along the length of the virtual artery (2000 μm).

Electron microscopy

Electron microscopy was conducted according to standard procedures (Sandow *et al.* 2002, 2004). In brief, anaesthetized rats were perfused via the left ventricle with saline (0.9% NaCl) containing 0.1% $NaNO_3$, 0.1% bovine serum albumin and 5 U ml^{-1} heparin at 25°C. Once cleared of blood, animals were fixed (10 min, 25°C) in 3% glutaraldehyde and 1% paraformaldehyde in 0.1 mM sodium cacodylate buffer with 0.2 mM $CaCl_2$, 15 mM sucrose and 10 mM betaine (pH 7.35). Resistance arteries were then: (1) dissected free of surrounding tissue; (2) postfixed in a 2% osmium tetroxide buffer (2 h); (3) stained with saturated uranyl acetate (2 h); and (4) embedded in Araldite 502 according to conventional procedures (Sandow *et al.* 2002, 2004). Arteries were transversally sectioned (100 nm thick) and prepared for transmission electron microscopy (TEM). Selected areas were viewed on a Hitachi 7100 TEM and photographed on plate film.

Chemicals, drugs and enzymes

Acetylcholine, $BaCl_2$, bradykinin, buffer reagents, collagenases (types F and H), indomethacin, N^{Ω}-nitro-L-arginine methyl ester (L-NAME), ouabain, phenylephrine, pinacidil and UTP were purchased from Sigma-Aldrich. Papain and P-1075 were acquired from Worthington (Lakewood, NJ, USA) and TOCRIS (Northfield, UK), respectively. When required, agents were dissolved in DMSO (final solvent concentration did not exceed 0.05%). All electron microscopy reagents were obtained from Electron Microscopy Sciences (Hatfield, PA, USA).

Statistical analysis

Data are expressed as means ± S.E.M. and n indicates the number of vessels or cells. No more than two experiments were performed on vessels or cells from a given animal. Paired t tests were performed to compare the effects of a given condition/treatment on arterial diameter, V_M or whole-cell current. $P = 0.05$ was considered statistically significant.

Results

K_{IR} expression in vascular smooth muscle

In this study we characterized the diversity of smooth muscle K_{IR} channel properties in middle cerebral, coronary septal and third-order mesenteric arteries. We began by monitoring the ability of preconstricted resistance arteries, stripped of endothelium, to respond to elevated extracellular [K^+] (from 5.8 to 15.8 mM), which augments K_{IR} channel activity. Arteries were preconstricted with elevated intravascular pressure and superfusion of phenylephrine in order to induce a depolarization that minimizes K_{IR} activity under basal conditions. Elevated [K^+] elicited a sustained Ba^{2+}-sensitive dilatation in cerebral and coronary arteries (Fig. 1). No similar response

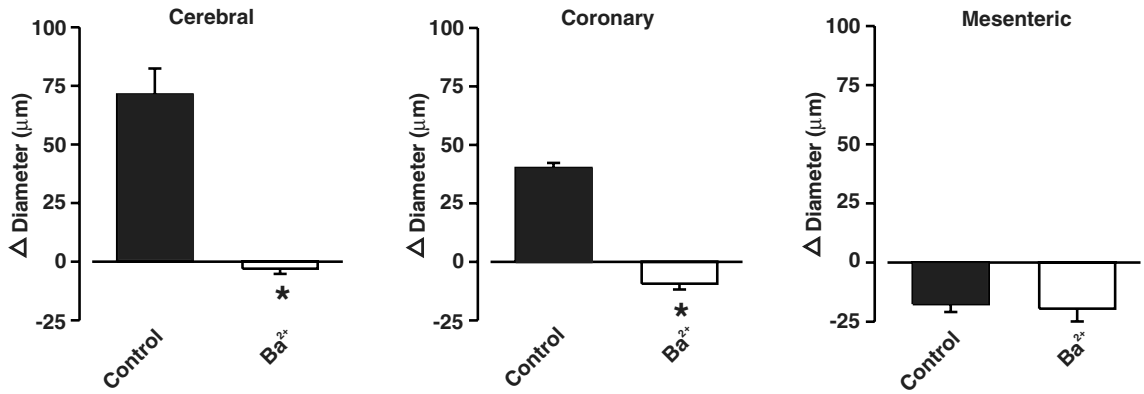

Figure 1. K_{IR} channels in cerebral and coronary arteries
Middle cerebral ($n = 6$), coronary septal ($n = 5$) and third-order mesenteric ($n = 6$) arteries were cannulated and preconstricted with elevated intravascular pressure (80 mmHg) and superfusion of phenylephrine (0.01–1 μM). Extracellular [K^+] was elevated from 5.8 to 15.8 mM for 5 min and the sustained vasomotor response was monitored in the absence and presence of Ba^{2+} (30 μM). Arterial diameter under control conditions, in the presence of Ba^{2+} and in Ca^{2+}-free media were as follows (in μm): cerebral, 120 ± 9, 115 ± 11, 281 ± 12; coronary, 123 ± 15, 117 ± 14, 199 ± 11; mesenteric, 134 ± 7, 127 ± 12, 278 ± 19.8, respectively. *Significantly different from the control response.

was present in mesenteric arteries, which is suggestive of differential K_{IR} expression. To better ascertain the variability in smooth muscle K_{IR} expression, a range of molecular and electrophysiological approaches were subsequently employed. We first quantified the relative mRNA expression of $K_{IR}2.x$ subunits in smooth muscle cells isolated from the three vessel types. Figure 2 shows that KIR2.1 and 2.2 mRNA were present in all smooth muscle aliquots although the relative expression was ∼2- to 6-fold higher in samples from cerebral and coronary arteries compared to mesenteric vessels. mRNA for $K_{IR}2.3$ and 2.4 was absent from all smooth muscle aliquots. Western blot analysis confirmed the presence of $K_{IR}2.1/2.2$ protein in whole arteries (data not shown). We did not, however, use this approach for cell-specific quantification because whole arteries contain several cell types that express K_{IR} subunits. We also avoided immunocytochemistry given its limited quantitative power. Instead, patch-clamp electrophysiology was used to monitor the magnitude of the Ba^{2+}-sensitive inward current. Figure 3 reveals that a Ba^{2+}-sensitive inward current was prominent in smooth muscle cells from cerebral and coronary arteries. No similar current was present in mesenteric smooth muscle cells, which is consistent with low K_{IR} expression. Despite this absence, inward current was sizable in mesenteric smooth muscle cells. The ionic basis of this current(s) remains unclear and warrants future investigation.

It is important to note that whereas K_{IR} expression is low in mesenteric arteries, these arteries can dilate in response to elevated extracellular $[K^+]$ if the protocol is designed to stimulate the Na^+–K^+ pump. For example, if resting extracellular $[K^+]$ was first reduced from 5.8 to 4 mM to decrease basal Na^+–K^+-ATPase activity, the subsequent elevation of extracellular $[K^+]$ did elicit a transient dilatation (Fig. 4*A*). This response often displayed dilatation and was absent after the application of ouabain but not Ba^{2+} (Fig. 4*B* and *C*).

Defining negative slope conductance

With negative slope conductance proposed to be an important K_{IR} property, we focused on documenting this biophysical characteristic. Vascular smooth muscle cells were enzymatically isolated and whole-cell patch-clamp electrophysiology used to quantify the outward component of the K_{IR} current. Using bath and pipette solutions that facilitate current flow, a Ba^{2+}-substracted outward current, greater than 1.5 pA, was observed in smooth muscle cells isolated from cerebral (4 of 8; peak outward current, 4.7 ± 0.8 pA at -24.8 ± 2.6 mV) and coronary (3 of 6; peak outward current, 1.9 ± 0.7 pA at -15.0 ± 1.8 mV) arteries (Fig. 5). Although small in magnitude, negative slope conductance was readily observed between −10 and +10 mV. By contrast, outward current was not discernable in mesenteric smooth muscle cells (0 of 6). If negative slope conductance is important in arterial V_M regulation, then micromolar levels of Ba^{2+} should strongly depolarize arteries maintained at more negative potentials. Consistent with this prediction, the results in Fig. 6 illustrate that cerebral and coronary arteries stripped of endothelium and sustained in a hyperpolarized state (by lowering intravascular pressure) depolarized in response to superfused Ba^{2+} (cerebral, 52.4 ± 1.2 to -36.2 ± 1.0 mV; coronary, -50.6 ± 1.4 to -35.2 ± 1.0 mV). When depolarized by high intravascular pressure and superfused phenylephrine, Ba^{2+}-induced responses were reduced to < 2 mV (cerebral, -36.4 ± 2.1 to -35.4 ± 2.0 mV; coronary, -35.0 ± 0.5 to -34.1 ± 0.5 mV). Ba^{2+}-induced depolarization was negligible in mesenteric arteries stripped of endothelium and maintained in a

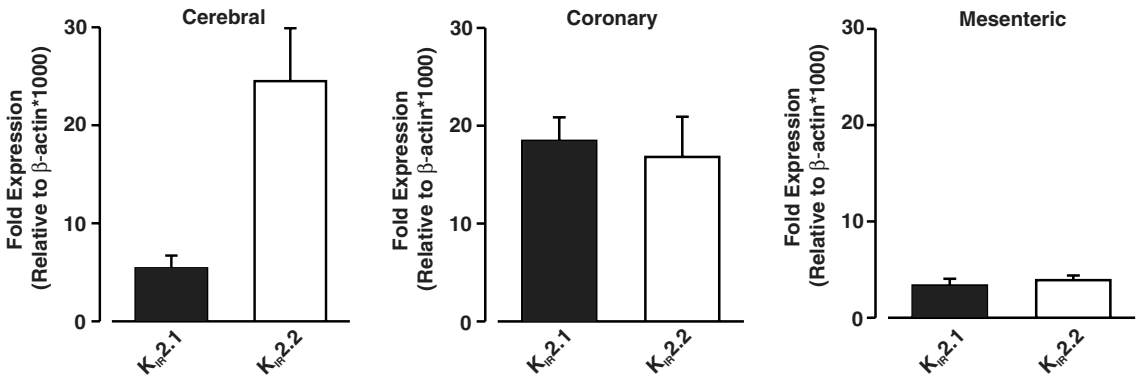

Figure 2. Real-time PCR analysis of $K_{IR}2$ subtypes
mRNA expression of $K_{IR}2.1$ and $K_{IR}2.2$ in smooth muscle cells (∼200) enzymatically isolated from middle cerebral ($n = 8$), coronary septal ($n = 7$) and third-order mesenteric ($n = 8$) arteries. Data are expressed relative to β-actin × 1000. Note that KIR2.3 and $K_{IR}2.4$ mRNA was not detected in smooth muscle samples free of endothelial contamination.

hyperpolarized state by low intravascular pressure (-57.2 ± 0.9 to -55.8 ± 1.1 mV).

Exploring electrical amplification

With knowledge of K_{IR} expression and negative slope conductance, next we investigated whether these channels amplify responses initiated by other K^+ conductances. We began at a theoretical level with a computational model designed to mimic a multilayered artery. Under control conditions, the injection of 4.6 pA hyperpolarizing current into each smooth muscle cell reduced V_M from -40 mV to approximately -55 mV (Fig. 7). Consistent with the amplification concept, the removal of K_{IR} from the smooth muscle ionic representation attenuated hyperpolarization. To support these theoretical results, we examined whether Ba^{2+} application altered the ability of middle cerebral, coronary septal and third-order mesenteric arteries to respond to K^+ channel activators. The results in Fig. 8 illustrate that 30 μM Ba^{2+} does attenuate K_{ATP}-induced dilatation in arteries that were de-endothelialized and which express smooth muscle K_{IR} channels (i.e. cerebral and coronary). By contrast, Ba^{2+} had little effect on K_{ATP}-induced responses in mesenteric arteries where smooth muscle K_{IR} expression is limited. This unique pattern of Ba^{2+} attenuation extended to other agents including those that modulate endothelial K^+ channels. This was evident in the experiments of Fig. 9 with endothelial-intact vessels, where superfused Ba^{2+} limited the ability of cerebral and coronary, but not mesenteric arteries to dilate in response to intraluminal UTP or superfused acetylcholine. These agents are known to activate endothelial Ca^{2+}-activated K^+ channels (Marrelli *et al.* 2003; Crane *et al.* 2003*a*; Gluais *et al.* 2005; McNeish *et al.* 2006). Note that intraluminal UTP was employed in the cerebrovascular experiments because superfused acetylcholine did not elicit a significant endothelium-dependent response. The preceding experiments were conducted in the presence of L-NAME (50 μM) and indomethacin (10 μM) to limit nitric oxide and prostaglandin production, respectively. The inherent ability of the endothelium to hyperpolarize the overlying smooth muscle could arise from the expression of myo-endothelial gap junctions and the passage of charge between the two cell layers. As shown in Fig. 10, myo-endothelial contact sites were prevalent in all three vessel types. These structures formed at the tip of endothelial projections which penetrated through the

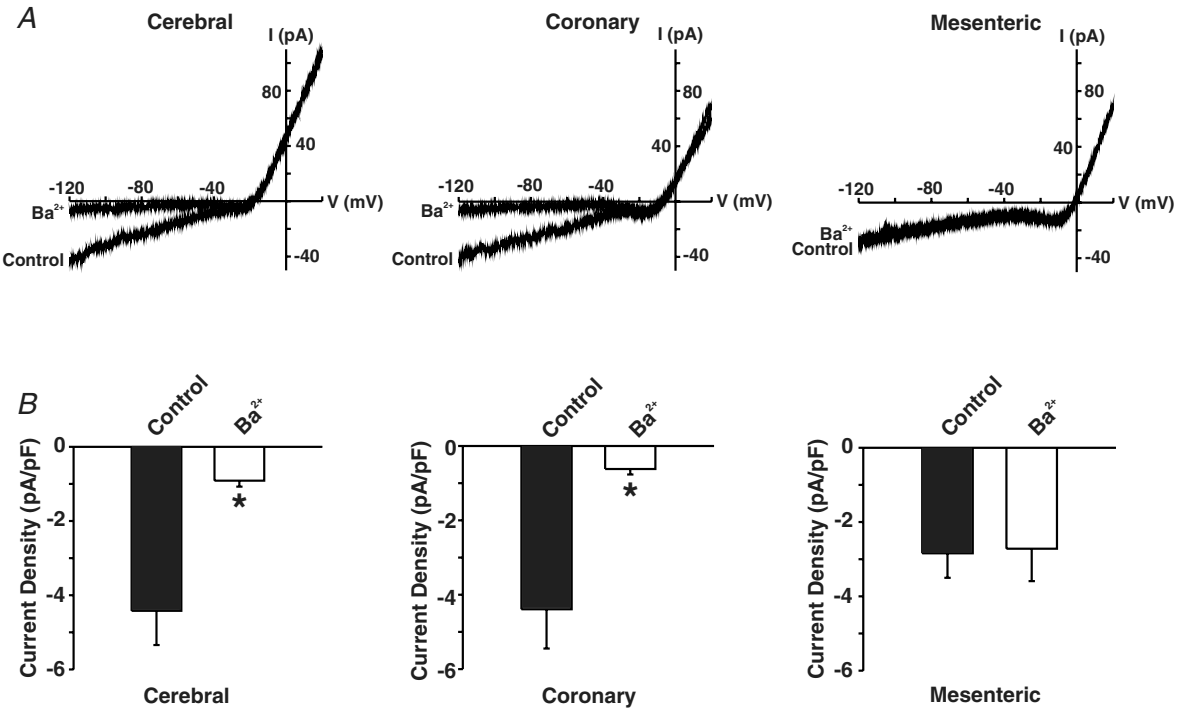

Figure 3. The Ba^{2+}-sensitive K_{IR} current in vascular smooth muscle cells
Whole-cell currents were monitored in middle cerebral, coronary septal and third-order mesenteric smooth muscle cells in the absence and presence of Ba^{2+} (30 μM). The intra- and extracellular $[K^+]$ were 120 and 60 mM, respectively. *A*, representative whole-cell current in the absence and presence of Ba^{2+}. *B*, peak inward current (at -120 mV) in cerebral ($n = 8$), coronary ($n = 6$) and mesenteric ($n = 6$) smooth muscle cells prior to and following Ba^{2+} application. *Significantly different from control.

internal elastic lamina. High-resolution electron microscopy demonstrated gap junctional plaques at these sites of myo-endothelial contact.

Discussion

In this study we examined whether smooth muscle K_{IR} channels amplify arterial responses initiated by smooth muscle or endothelial K^+ conductances. To accomplish this objective it was necessary to: (1) characterize K_{IR} expression in selected resistance arteries; (2) demonstrate negative slope conductance; and (3) determine whether K_{IR} inhibition attenuates vessel responsiveness. Using a range of functional, molecular and electrophysiological approaches, initial observations revealed that smooth muscle K_{IR} channels are highly expressed in middle cerebral and septal coronary but not third-order mesenteric arteries. When present, smooth muscle K_{IR} channels were composed of $K_{IR}2.1$ and 2.2 subunits and displayed negative slope conductance, which is a key biophysical property that ensures that channel activity increases with hyperpolarization. Computational simulations subsequently revealed that negative slope conductance could amplify the hyperpolarization initiated by another vascular K^+ conductance. This prediction was verified experimentally in cerebral and coronary arteries where 30 μM Ba^{2+}, a K_{IR} inhibitor, attenuated the ability of these vessels to dilate in response to smooth muscle and endothelial K^+ channel activators. Ba^{2+}-induced attenuation was absent in mesenteric arteries, which are vessels with low K_{IR} expression. Cumulatively, our findings demonstrate that when smooth muscle K_{IR} channels are expressed, this conductance can function as an electrical amplifier.

K_{IR} expression in vascular smooth muscle

K_{IR} channels are aptly named for their distinctive ability to pass current more readily in the inward direction. Structurally, these channels consist of four α-subunits,

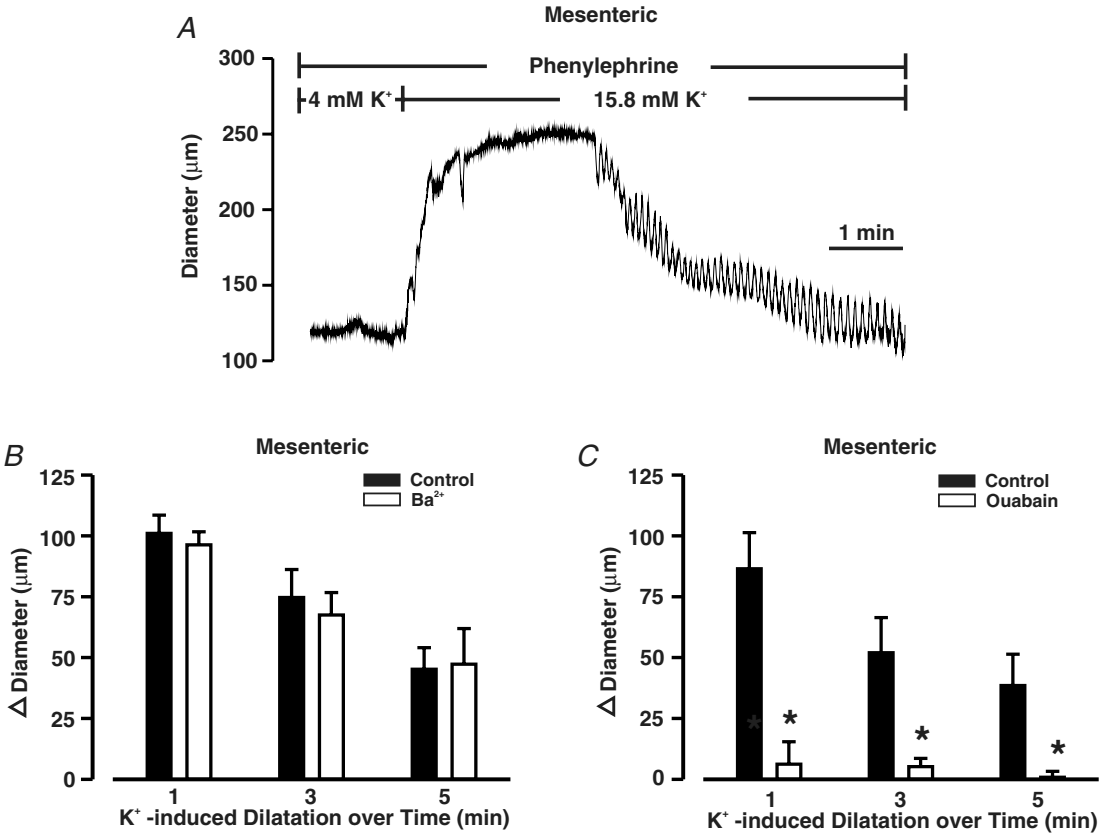

Figure 4. Na^+–K^+-ATPase activation initiates transient dilatation in mesenteric arteries
Third-order mesenteric arteries were cannulated and preconstricted with elevated intravascular pressure (80 mmHg) and superfusion of phenylephrine (~1 μM). Following equilibration, extracellular [K^+] was reduced from 5.8 to 4.0 mM to decrease Na^+–K^+-ATPase activity. Extracellular [K^+] was subsequently elevated to 15.8 mM eliciting a transient dilatation (*A*). This response was monitored in the absence and presence of Ba^{2+} (*A*; 30 μM; $n = 6$) or ouabain (*B*; 100 nM; $n = 6$). Arterial diameter in 4 mM K^+, 4 mM K^+ + Ba^{2+} or ouabain and in Ca^{2+}-free media were (in μm): Fig. 5*A*; 121 ± 8, 109 ± 7, 277 ± 17; Fig. 5*B*; 111 ± 13, 89 ± 13, 259 ± 16. *Significantly different from the control response.

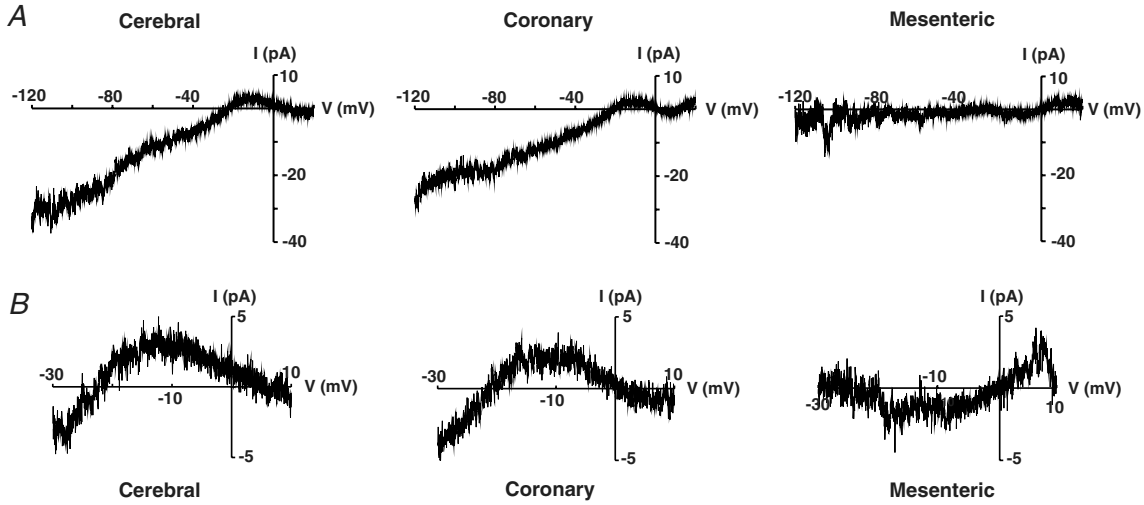

Figure 5. Outward current through smooth muscle K_{IR} channels
Whole-cell currents were monitored in middle cerebral, coronary septal and third-order mesenteric smooth muscle cells in the absence and presence of Ba^{2+} (30 μM). The intra- and extracellular [K^+] were 120 and 60 mM, respectively. The outward component in *A* was magnified and replotted in *B*. Note that a small outward current, greater than 2 pA and with negative slope conductance was detectable in cerebral (4 of 8) and coronary (3 of 6) smooth muscle cells. No similar current was observed in mesenteric smooth muscle cells (0 of 6).

each composed of two transmembrane domains and a GYG-containing 'P loop' that confers selective K^+ permeability (Quayle *et al.* 1997; Bichet *et al.* 2003). Molecular approaches have identified seven K_{IR} subfamilies, with K_{IR} 2.x channels readily expressed in excitable cells (Liu *et al.* 2001; Dhamoon *et al.* 2004; Wu *et al.* 2007). This subfamily is composed of four members with KIR2.1 and 2.2 being present in vascular smooth muscle (Bradley *et al.* 1999; Karkanis *et al.* 2003; Wu *et al.* 2007). Although it is clear that smooth muscle K_{IR} channels are present in cerebral and coronary arteries (Quayle *et al.* 1993, 1996; Robertson *et al.* 1996), expression may not be universal to all vascular beds (Crane *et al.* 2003*b*). Indeed, past studies have implied, from limited observations, that smooth muscle K_{IR} channels are poorly expressed in the mesenteric circulation (Crane *et al.* 2003*b*; Brochet & Langton, 2006). To better understand channel expression patterns, we characterized the properties of smooth muscle K_{IR} channels in middle cerebral, coronary septal and third-order mesenteric arteries. This characterization began at a functional level where elevated extracellular [K^+] (5.8–15.8 mM) was used to augment K_{IR} activity and dilate resistance arteries preconstricted with elevated intravascular pressure and superfusion of phenylephrine.

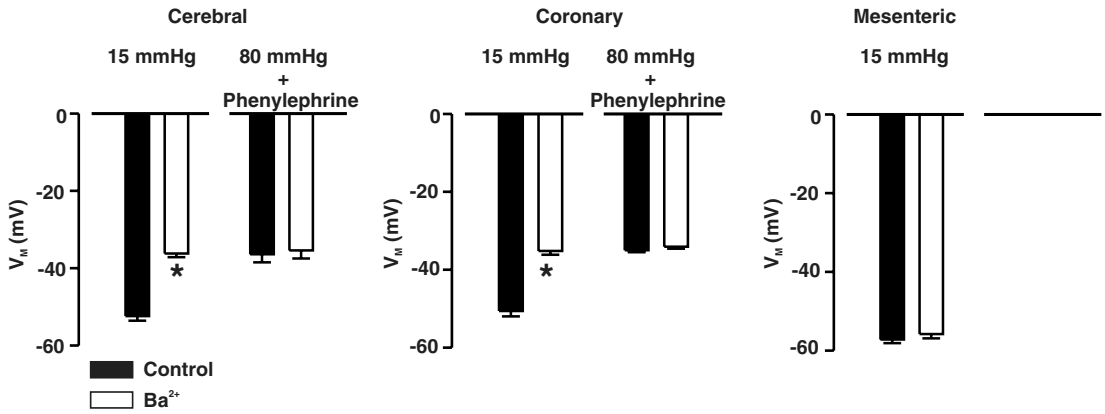

Figure 6. The effects of Ba^{2+} on arterial membrane potential (V_M)
Middle cerebral ($n = 5$), coronary septal ($n = 6$) and third-order mesenteric ($n = 5$) arteries were cannulated and maintained in either a hyperpolarized (intravascular pressure, 15 mmHg) or depolarized (intravascular pressure, 80 mmHg; superfused phenylephrine, 0.01–1 μM) state. Arterial V_M was monitored in the absence and presence of Ba^{2+} (30 μM). *Significantly different from control V_M.

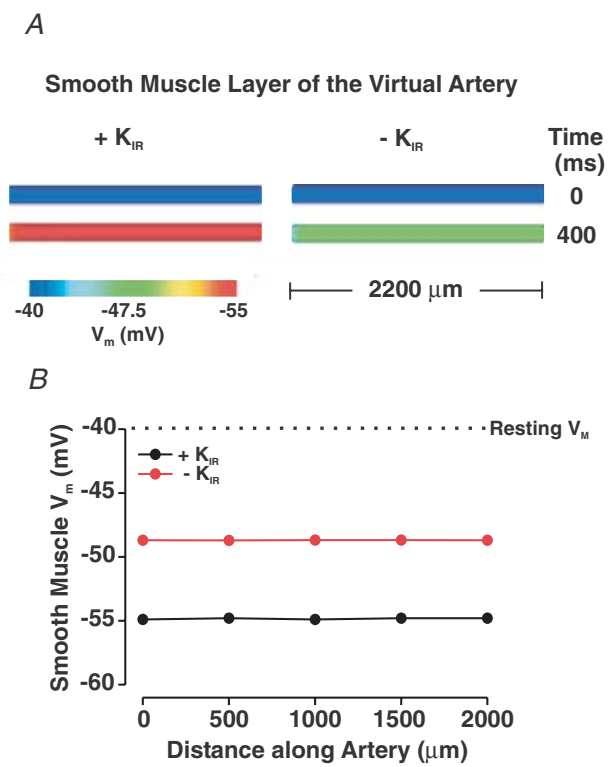

Figure 7. Computational modelling predicts that K_{IR} channels amplify hyperpolarization
Simulation: a hyperpolarizing current was injected (4.6 pA for 500 ms) into each smooth muscle cell of a virtual artery composed of one endothelial layer and four smooth muscle layers. Membrane potential responses were monitored (at 500 ms) in the outer smooth muscle cell layer in the presence and absence of a smooth muscle K_{IR}-like current. These responses were colour-mapped along the vessel wall (*A*) or were presented in a two-dimensional voltage plot (*B*).

As is evident in Fig. 1, this perturbation elicited a differential response with cerebral and coronary arteries dilating in a Ba^{2+}-sensitive manner, but with mesenteric vessels failing to respond similarly. With these functional observations implying variable K_{IR} expression, we subsequently isolated smooth muscle cells and utilized real-time PCR to quantify mRNA expression. This analysis revealed that whereas KIR2.1/2.2 mRNA was present in all vascular beds, expression levels were 2- to 6-fold higher in smooth muscle cells from cerebral and coronary arteries than those from the mesenteric circulation (Fig. 2). Whereas Western blot analysis verified K_{IR} protein expression (data not shown), the mixed nature of a whole vessel renders this approach inappropriate to quantify differential channel expression. Immunocytochemistry was also avoided due its limited quantitative power. Instead, the magnitude of the Ba^{2+}-sensitive inward current was examined in smooth muscle cells isolated from the three vascular beds. Consistent with our functional and molecular observations, a Ba^{2+}-sensitive inward current was detected in smooth muscle cells from cerebral and coronary arteries (Fig. 3). The magnitude of this current was comparable to that previously observed (Quayle *et al.* 1993, 1996; Robertson *et al.* 1996) and was not present in mesenteric smooth muscle cells. Overall, this work is the first to illustrate a consistent pattern of greater smooth muscle K_{IR} expression in cerebral and coronary arteries than in the mesenteric circulation.

Although K_{IR} expression is low in mesenteric smooth muscle cells, it should be stated that this absence does not preclude these vessels from dilating in response to certain K^+ challenges. Indeed, as highlighted in Fig. 4, mesenteric arteries can dilate in response to 15.8 mM K^+ if resting extracellular $[K^+]$ is reduced to 4.0 mM prior to stimulation. This reduction in resting extracellular $[K^+]$ presumably enables mesenteric dilatation via limited basal Na^+–K^+-ATPase activity, thereby making the pump available for subsequent stimulation (Brochet & Langton, 2006). This view is consistent with the transient nature of the dilatory response along with its documented ouabain sensitivity (Fig. 4).

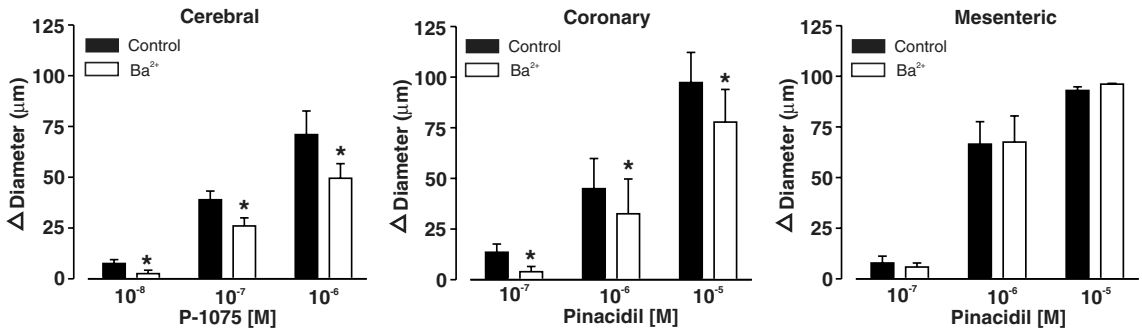

Figure 8. The effects of Ba^{2+} on K_{ATP}-induced vasodilatation
Middle cerebral ($n = 6$), coronary septal ($n = 6$) and third-order mesenteric ($n = 6$) arteries were cannulated and preconstricted with elevated intravascular pressure (80 mmHg) and superfusion of phenylephrine (0.01–1 μM). Vasodilatory responses to K_{ATP} openers (P-1075 or pinacidil) were monitored in the absence or presence of Ba^{2+} (30 μM). Arterial diameter under control conditions, in the presence of Ba^{2+} and in Ca^{2+}-free media were as follows (in μm): cerebral, 125 ± 7, 115 ± 8, 249 ± 7; coronary, 137 ± 21, 135 ± 16, 262 ± 29; mesenteric, 77 ± 5, 76 ± 5, 256 ± 18, respectively. *Significantly different control K_{ATP}-induced dilatation.

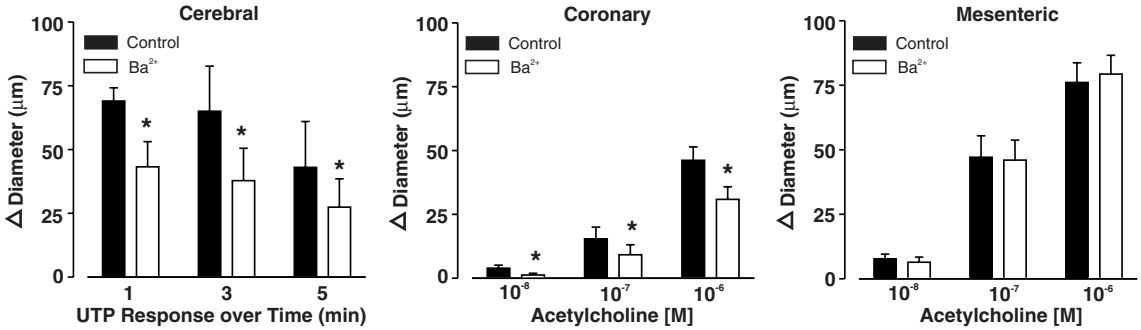

Figure 9. Effects of Ba^{2+} on endothelium-induced vasodilatation
Middle cerebral ($n = 6$), coronary septal ($n = 6$) and third-order mesenteric ($n = 6$) arteries were cannulated and preconstricted with elevated intravascular pressure (80 mmHg) and superfusion of phenylephrine (0.01–1 μM). L-NAME (100 μM) and indomethacin (10 μM) were added to attenuate nitric oxide and prostaglandin production, respectively. Dilatory responses to intraluminal UTP (cerebral) or superfused acetylcholine (coronary and mesenteric) were monitored in the absence or presence of Ba^{2+} (30 μM). Arterial diameter under control conditions, in the presence of Ba^{2+} and in Ca^{2+}-free media were as follows (in μm): cerebral, 214 ± 10, 204 ± 9, 302 ± 8; coronary, 146 ± 17, 135 ± 14, 269 ± 17; mesenteric 89 ± 7, 87 ± 5, 247 ± 19, respectively. *Significantly different control dilatation.

Defining negative slope conductance

Although strong inward rectification is one of the defining characteristics of smooth muscle K_{IR} channels, it is the small outward component with its intrinsic property of negative slope conductance which may be important for the regulation of arterial tone. As such, in this study we focused on documenting this biophysical property. In the

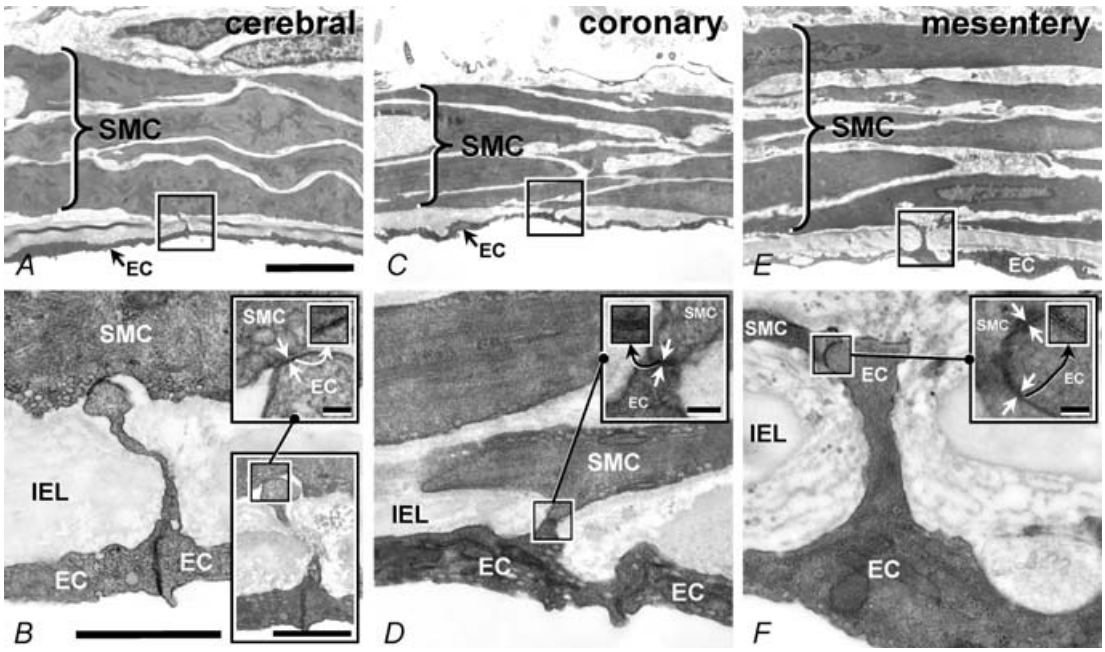

Figure 10. Myo-endothelial gap junctions in the arterial wall
Middle cerebral (*A* and *D*) coronary septal (*B* and *E*) and third-order mesenteric (*C* and *F*) arteries were isolated and prepared for electron microscopy. Myo-endothelial gap junctions with their characteristic pentalaminar appearance were common in these vessels; gap junctional plaques between endothelial cells were also evident (*B* and *D*; between arrowheads). Area within square in *A*, *C* and *E* corresponds to the main panels in *B*, *D* and *F*, respectively. Insets in *B*, *D* and *F* are shown in upper right of each respective panel. In *B* (main), the endothelial 'stalk' makes contact with an adjacent smooth muscle cell, while two serial sections later (lower inset), the myo-endothelial contact site reveals its typical pentalaminar appearance (upper inset). Middle cerebral, coronary septal and third-order mesenteric arteries had 3.8 ± 0.2, 4.2 ± 0.2 and 4.8 ± 0.2 smooth muscle cell layers, respectively ($n = 3$). Scale bar: *A*, *C* and *E*, 5 μm; *B*, *D* and *F* (and *B*, lower inset), 1 μm; *B*, *D* and *F* (upper inset), 50 nm.

past, such documentation has proven difficult because outward current is limited and close to the resolution limits of a patch-clamp amplifier (Quayle *et al.* 1993, 1996; Robertson *et al.* 1996; Wu *et al.* 2007). However, by carefully designing the bath and pipette solutions to augment K_{IR} activity, an outward current, greater than 2 pA and with negative slope conductance was resolvable in 50% of the smooth muscle cells from cerebral and coronary arteries (Fig. 5). Peak outward current did not exceed 5 pA and, as expected, a similar current was not apparent in mesenteric smooth muscle cells. Although the preceding single-cell results are important, they alone do not demonstrate that this biophysical property is present in intact arteries. To address this concern, we measured arterial V_M and the effects of Ba^{2+} on vessels maintained at different resting potentials through manipulation of intravascular pressure and superfusion of phenylephrine. The logic underlying this approach centres on the idea that if negative slope conductance is present, then Ba^{2+} should have a greater electrical effect on arteries held at more negative potentials. Consistent with its functional occurrence, micromolar levels of Ba^{2+} strongly depolarized (∼15 mV) K_{IR}-expressing arteries maintained in a hyperpolarized state by low intravascular pressure (Fig. 6). At more depolarized potentials where outward K_{IR} current should be limited, Ba^{2+}-induced depolarization was restricted to < 2 mV. In mesenteric arteries where smooth muscle K_{IR} expression was low, Ba^{2+} application had little discernable effect on resting V_M. This latter experiment is an important control as it demonstrates that at 30 μM, Ba^{2+} does not substantively affect other channels involved in V_M regulation. This is an important consideration because at submillimolar concentrations, this divalent cation can attenuate the activity of K_{ATP} and twin-pore K^+ channels (Nelson *et al.* 1990; Nelson & Quayle, 1995; Campanucci *et al.* 2003).

Exploring electrical amplification

Lastly, with knowledge of vascular smooth muscle K_{IR} properties, we focused on whether these channels amplify responses initiated by other smooth muscle or endothelial K^+ conductances. As stated previously, our interest in this channel stems from the intrinsic property of negative slope conductance. Briefly, negative slope conductance refers to the inherent ability of K_{IR} channels to increase their activity as a cell hyperpolarizes (Nelson *et al.* 1990; Nelson & Quayle, 1995). This rise depends on the relief of a voltage-dependent Mg^{2+}/polyamine block (Nelson & Quayle, 1995; Robertson *et al.* 1996; Matsuda *et al.* 2003). Our work began at a theoretical level with the use of a computational model to explore whether a smooth muscle K_{IR} current alters the ability of a multilayered artery to hyperpolarize (Fig. 7). The K_{IR} current employed in this model retained negative slope conductance and was designed to be minimally and maximally active at −40 and −60 mV, respectively. Peak outward current was set to 2 pA, a value consistent with a limited set of observations gathered at physiological $[K^+]$ (Wu *et al.* 2007). In keeping with the amplification concept, the hyperpolarization induced by smooth muscle current injection decreased in the absence of K_{IR} (Fig. 7). This attenuation approached 35% and a change of this magnitude should, under experimental conditions, impair arterial dilatation. To verify this prediction, we examined the ability of K_{IR} expressing (cerebral/coronary) and nominally expressing (mesenteric) arteries to dilate in response to K^+ channel activators in the presence and absence of 30 μM Ba^{2+}. In these experiments, all arteries were preconstricted with elevated intravascular pressure and superfusion of phenylephrine, which will induce depolarization and minimize K_{IR} activity under basal conditions. Consistent with theoretical predictions, initial experiments with de-endothelialized vessels reveal that Ba^{2+} attenuated the ability of presumed K_{ATP} activators to dilate cerebral and coronary arteries (Fig. 8). This reduction occurred over a range of concentrations and was absent in mesenteric arteries, which a show low K_{IR} expression. A similar pattern of attenuation was observed in a second experimental set where the endothelium was retained and intraluminal UTP or superfused acetylcholine was used to activate small- and/or intermediate-conductance Ca^{2+}-activated K^+ channels (Fig. 9) (Marrelli *et al.* 2003; Crane *et al.* 2003*a*; Gluais *et al.* 2005; McNeish *et al.* 2006). It is interesting to note that L-NAME and indomethacin were included in the latter experiments. As such, it can be argued that the endothelium is driving the smooth muscle responses through a nitric oxide- and prostaglandin-independent event. The nature of this event is unclear but could involve the production of an arachidonic acid metabolite (Campbell *et al.* 1996; Bryan *et al.* 2006; Marrelli *et al.* 2007) or perhaps hydrogen peroxide (Hatoum *et al.* 2005). Alternatively, charge could directly spread from the endothelium to the overlying smooth muscle layers via myo-endothelial gap junctions (Emerson & Segal, 2000; Sandow *et al.* 2002; Sokoya *et al.* 2007). Previous studies have provided both direct and indirect evidence for these junctions in cerebral, coronary and mesenteric arteries (Edwards *et al.* 1998; Edwards *et al.* 2000; Mather *et al.* 2006). Consistent with these observations and the concept of direct charge transfer, high-resolution electron microscopy illustrated that myo-endothelial gap junctions were prevalent in all three arteries (Fig. 10). These junctions occur at the tip of endothelial cell projections which pass through the internal elastic lamina to make contact with the overlying smooth muscle.

As a final note, it is important to recognize that while this study is the first to associate negative slope conductance

with the facilitation of endothelium-dependent responses, it is not the first to implicate smooth muscle K_{IR} channels in such a process (Edwards *et al.* 1998; Edwards & Weston, 2004). Indeed, Edwards & Weston (2004) have previously drawn this connection, although unlike in this investigation, negative slope conductance was not considered. Rather, these authors proposed that K_{IR} channels were directly activated by the K^+ release from endothelial cells (Edwards *et al.* 1998; Edwards & Weston, 2004). This is an interesting hypothesis although one that is difficult to resolve given that the relative area of the endothelium is small compared to the extracellular space. As such, it would be difficult to sustain a millimolar rise in extracellular $[K^+]$ without inducing a substantial depletion of intracellular $[K^+]$. Such a depletion would elicit a rightward shift in the K^+ equilibrium potential and in a counterproductive manner limit arterial hyperpolarization.

Limitations

We acknowledge three important concerns in the manner in which the preceding observations are interpreted. First, we recognize that K_{IR} channels may be present in endothelial cells (Nilius & Droogmans, 2001; Fang *et al.* 2005), and thus part of the Ba^{2+}-induced attenuation observed in Fig. 9 could result from their inhibition. Second, we are aware that not all arteries need to express smooth muscle K_{IR} channels to elicit robust dilatory responses. This was evident from the mesenteric observations and it raises a series of intriguing biophysical questions for further investigation. Finally, we acknowledge that in addition to amplifying hyperpolarization, negative slope conductance could facilitate depolarization. Current experimental approaches do not, however, allow this possibility to be systematically tested.

Summary

The results of this study demonstrate that smooth muscle K_{IR} channels, when present, play an important role in amplifying electrical and vasomotor responses initiated by vascular K^+ channels. Amplification was ascribed to negative slope conductance, a biophysical property which enables KIR2.1/2.2 activity to increase with hyperpolarization. Broadly speaking, our findings illustrate that subtle channel properties are important for the regulation of arterial responsiveness. They also show that when an agonist alters the electrical state of a multilayered artery, the ion channels underlying this response are not always under the direct regulatory control of the stimulus.

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